Progesterone Inhibits Basal and Gonadotropin-Releasing Hormone Induction of Luteinizing Hormone β-Subunit Gene Expression

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LH and FSH play critical roles in mammalian reproduction by mediating steroidogenesis and gametogenesis in the gonad. Gonadal steroid hormone feedback to the hypothalamus and pituitary influences production of the gonadotropins. We previously demonstrated that progesterone differentially regulates the expression of the LH and FSH β-subunits at the level of the gonadotrope: FSHβ transcription is induced, whereas LHβ is repressed. In this study, we investigated the mechanism of progesterone repression of LHβ gene expression using immortalized gonadotrope-derived LβT2 cells. The progesterone suppression of both basal and GnRH-induced LHβ gene expression occurs in a hormone- and receptor-dependent manner. Chromatin immunoprecipitation demonstrates that the hormone-bound progesterone receptor (PR) is recruited to the endogenous mouse LHβ promoter. In addition, suppression requires both the amino-terminal and DNA-binding regions of PR. Furthermore, progesterone suppression does not require direct PR binding to the promoter, and, thus, PR is likely recruited to the promoter via indirect binding through other transcription factors. These data demonstrate that the molecular mechanism for progesterone action on the LHβ promoter is distinct from FSHβ, which involves direct PR binding to the promoter to produce activation. It also differs from androgen repression of LHβ gene expression in that, rather than Sp1 or steroidogenic factor-1 elements, it requires elements within −300/−250 and −200/−150 that also contribute to basal expression of the LHβ promoter. Altogether, our data indicate that progesterone feedback at the level of the pituitary gonadotrope is likely to play a key role in differential production of the gonadotropin genes. (Endocrinology 150: 2395–2403, 2009)

The mammalian hypothalamic-pituitary-gonadal axis controls reproduction, including sexual development, puberty, the menstrual cycle, pregnancy, and menopause. GnRH is secreted in a pulsatile manner directly into the hypophyseal portal system by neurons in the hypothalamus (1). GnRH then activates its receptor on the surface of gonadotrope cells within the anterior pituitary. Activation of the GnRH receptor leads to synthesis and secretion of LH and FSH, consisting of a common α-subunit and a unique β-subunit (2). These glycoprotein hormones are secreted into the bloodstream to regulate gametogenesis; folliculogenesis; and production of testosterone, estrogen, and progesterone in the gonads (3–5). Subsequently these steroids feed back to regulate expression and secretion of GnRH, LH, and FSH in the hypothalamus and the anterior pituitary.

Because synthesis of LHβ and FSHβ is the rate-limiting step in gonadotropin production (6, 7), transcriptional regulation is a key focus. The conserved proximal promoter of LHβ binds several transcriptional regulators that modulate cell-specific expression including steroidogenic factor (SF)-1 (8, 9), early growth response protein (Egr)-1, and pituitary homeobox 1/orthodenticle homeobox 1 (10–12). These proteins, as well as those binding to more distal elements, interact to elicit basal promoter activity. GnRH induction of the rat LHβ promoter has been shown to involve a tripartite GnRH response element, com-

Abbreviations: AR, Androgen receptor; ChIP, chromatin Immunoprecipitation; DBD, DNA-binding domain; Egr, early growth response protein; PR, progesterone receptor; PRE, progesterone response element; SF, steroidogenic factor; TK, thymidine kinase.
posed of two Sp1 sites in the distal promoter, and proximal pairs of SF-1 and Egr-1 binding elements (10).

In addition to GnRH, steroid hormones play a pivotal role in LH synthesis. Many studies have shown that LHβ mRNA levels increase after gonadectomy and subsequently decrease with re-introduction of gonadal steroid hormones (13, 14). Steroid hormone receptors are expressed in gonadotrope cells (15–17), indicating that regulation by gonadal steroids likely occurs directly at the level of the pituitary, in addition to the hypothalamus. Estrogens suppress LHβ synthesis due to negative feedback at the hypothalamus (18, 19), but they also exert a direct effect on the pituitary by inducing LH gene expression in gonadotropes (20, 21). In addition, many studies indicate that androgens repress LHβ mRNA directly in the pituitary. Androgens decrease LHβ mRNA levels in castrated, GnRH antagonist-treated rats (22) and primary pituitary cells (23). Furthermore, androgens suppress LHβ gene expression in immortalized gonadotrope cells (24, 25).

Despite evidence for progesterone enhancement of GnRH-induced LH secretion (26), it is unclear whether progesterone regulates LHβ gene expression directly in the pituitary gonadotrope. Because levels of LHβ mRNA are lowest during the luteal phase of the ovulatory cycle when circulating progesterone levels are at their highest, progesterone is an excellent candidate for countering the stimulatory effects of GnRH on LHβ gene expression. Many studies have shown that, in the presence of estrogen, progesterone can suppress LHβ mRNA levels in rodents (27–29), although these experiments did not differentiate between hypothalamic and pituitary sites of action. Contrary to other studies that did not report an effect of progesterone on LHβ (30, 31), we recently demonstrated that progesterone can suppress LHβ gene expression in gonadotrope cells, indicating that progesterone may serve as a differential regulatory influence on the two gonadotropins during the menstrual cycle.

In mammals, gonadotropes comprise approximately 10% of the anterior pituitary cell population, making it difficult to conduct mechanistic studies of their function (33). Development of the immortalized LβT2 cells provided the opportunity to analyze mechanisms of steroid hormonal regulation of the gonadotropin genes in the context of a pure population of gonadotrope cells (34). The immortalized LβT2 cell line expresses many markers of a mature gonadotrope including FSHβ, LHβ, activin, follistatin, and inhibin as well as activin, inhibin, and steroid receptors (32, 35–38). In this study, we used transient transfection experiments in LβT2 cells to demonstrate that progesterone suppresses both basal transcription and GnRH-induced LHβ gene expression in gonadotrope cells. Because estrogen is necessary for the induction of progesterone receptor (PR) but may mask the repressive effects of progesterone due to its stimulatory effect on LHβ gene expression, we enhanced PR levels directly by overexpressing the receptor in LβT2 cells. We defined and characterized a progesterone-responsive region in the proximal LHβ promoter necessary and sufficient for the suppressive effect. We determined that progesterone suppression is dependent on the presence of PR and that PR is recruited to the endogenous LHβ promoter after hormone treatment in LβT2 cells. Interestingly, direct binding of PR to the LHβ promoter does not appear to be necessary for the suppression, although the DNA-binding domain (DBD) of PR is required. These results suggest that the progesterone suppression of LHβ gene expression is through indirect binding of PR to the DNA via other transcription factors.

Materials and Methods

Cell culture and transient transfection

Cell culture and transient transfections were performed as described previously (32). LβT2 cells were plated in 12-well plates at 3 × 10^5 cells/well. Wells were transfected with 400 ng of reporter plasmid and 100 ng of a PR expression vector, unless otherwise noted. Cells were also transfected with 200 ng of a thymidine kinase (TK)-β-gal reporter as a control for transfection efficiency. Six hours after transfection, cells were switched to serum-free DMEM. Eighteen hours later, cells were treated for 6 h (unless otherwise noted) with one of the following treatments: 0.1% ethanol and 0.1% BSA (vehicle control); 10^{-7} M promegestone (R5020) and 0.1% BSA; 0.1% ethanol and 10^{-7} M GnRH; or both 10^{-7} M R5020 and 10^{-8} M GnRH. R5020 was purchased from NEN Life Science Products Life Sciences (Boston, MA) and progesterone, mifepristone (RU486) and GnRH from Sigma (St. Louis, MO). Luciferase and β-galactosidase activity were measured as previously described (32). Transfection data were analyzed by one-way ANOVA, followed by post hoc comparisons with the Tukey-Kramer honestly significant difference test or two-way ANOVA using the statistical package JMP 7.0 (SAS, Cary, NC).

Subcloning and mutagenesis

PCR was performed using the appropriate primers (supplemental Table 1, published as supplemental data on The Endocrine Society’s Journals Online web site at http://endo.endojournals.org) to create 5′ truncations of the −1800-bp LHβ promoter at −500, −300, −150, and −87. Fragments were inserted between KpnI and HindIII in pGL3. A similar strategy was used to insert −300/−150 of the LHβ promoter into a −81-TK-luc reporter in forward or reverse orientation.

The PRB C577A DBD mutant unable to bind DNA was described previously (32). The QuikChange Kit (Stratagene, La Jolla, CA) was used to generate the 5′ truncations of the −1800-bp LHβ promoter at −500, −300, −150, and −87. Fragments were inserted between KpnI and HindIII in pGL3. A similar strategy was used to insert −300/−150 of the LHβ promoter into a −81-TK-luc reporter in forward or reverse orientation.

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EMSA

Full-length human PRB was overexpressed in S9 cells via a baculovirus expression system. The cell lysate was centrifuged at 40,000 rpm for 30 min, and the supernatant was taken as a soluble whole-cell extract or purified as described previously (39) and used in EMSA as described (32). The 1294 PR mouse monoclonal antibody was used to supershift PR and mouse IgG was used as a control. Oligonucleotides used for EMSA were shown in supplemental Table 1, published as supplemental data on The Endocrine Society’s Journals Online web site at http://endo.endojournals.org.

Chromatin immunoprecipitation (ChIP)

Confluent LβT2 cells in 15-cm plates were treated with vehicle or 10^{-7} M R5020 and cross-linked with 1% formaldehyde. The nuclear fraction was obtained and chromatin was sonicated. Protein-DNA complexes were incubated overnight with 1294 PR antibody or IgG control and precipitated with protein A/G beads. Beads were washed, the protein-DNA complexes eluted, the cross-links reversed, and the DNA precipitated, as described (32). PCR primers for the LHβ promoter spanned
the 220-bp sequence in the mouse LHβ gene from −180 to +40 (supplemental Table 1).

**Results**

**LHβ gene expression is suppressed in immortalized gonadotropes by progesterone**

Recently we demonstrated that both testosterone and progesterone can differentially modulate gonadotropin synthesis in pituitary gonadotrope cells by enhancing FSHβ and repressing LHβ gene expression (32). Because testosterone has been shown to suppress GnRH induction of LHβ gene transcription (24, 25), we hypothesized that progesterone might have a similar effect. Progesterone treatment suppressed both basal promoter activity and GnRH-induced LHβ gene transcription (Fig. 1A). Progesterone repressed basal gene expression by 10%. Treatment of the LβT2 cells with GnRH alone resulted in a 4.8-fold induction, which was reduced 65% by progesterone. This suppression was maintained for over 24 h (data not shown). Treatment of the cells with the synthetic progestin, R5020, elicited a similar suppression on basal and GnRH-induced LHβ gene expression (Fig. 1A), indicating that R5020 acts in an analogous manner to progesterone.

To test whether suppression of GnRH induction was dose-dependent, cells were treated with vehicle, 10−7 M R5020, 10−8 M GnRH, or 10−8 M GnRH and increasing concentrations of R5020 from 10−12 to 10−7 M (Fig. 1B). In these experiments, the progestin alone repressed basal LHβ gene expression by 30% (shown on an expanded scale as an inset in Fig. 1B). GnRH treatment resulted in a 4.2-fold induction. As the R5020 concentration was increased, the suppression of GnRH induction increased in concert until saturation at 10−9 M (approximately 70% reduction by 10−9 to 10−7 M). All subsequent experiments used 10−7 M R5020 to ensure saturation of the receptor. Together these experiments demonstrate that the progesterone regulation of LHβ occurs in a saturable, dose-dependent manner.

**Ligand-bound PR is necessary for maximal suppression of LHβ gene expression**

We next examined whether the suppression of LHβ by progesterone required the presence of its classical nuclear receptor. LβT2 cells were transfected with increasing concentrations of the PRB expression vector to mimic estrogen induction of PR levels without the complication of estrogen action on LHβ gene expression. Basal LHβ gene expression was not suppressed by R5020 unless PR was transfected into the cells (Fig. 1C). With 100 and 400 ng of PR expression vector, R5020 repressed basal gene expression by 25 and 33%, respectively. As increasing amounts of PR were transfected into the cells, GnRH induction (in the absence of R5020) decreased from 3.8-fold (when no receptor was added) to 3.5-fold with 100 ng PR and then down to 2.5-fold with 400 ng PR, indicating that the unliganded PR may slightly repress GnRH-induced LHβ gene expression (Fig. 1C), although it had no effect on basal LHβ levels (data not shown). Whereas GnRH induction decreased due to higher levels of exogenous receptor, the suppression of the GnRH induction by PR liganded with R5020 remained relatively constant. Thus, although low levels of PR are expressed endogenously (32, 40), transfection of exogenous receptor was necessary to observe significant suppression of both basal transcription and GnRH induction. Because suppression did not increase with greater amounts of exogenous receptor, all subsequent experiments used 100 ng of PR expression vector.

Once we demonstrated that the ligand-bound PR was required for suppression of LHβ gene expression by progesterone, we tested whether this effect could be blocked by the PR antagonist, RU486. Treatment with 100-fold higher levels of RU486 abrogated the suppressive effects of R5020 on basal and GnRH-induced LHβ gene expression (Fig. 1D). Interestingly, the GnRH induction was also suppressed 30% by the RU486 antagonist in the absence of the R5020 agonist.
forms are expressed in pituitary gonadotropes from the same gene and modulation of their ratio is thought to contribute to the degree of progesterone augmentation of GnRH-induced secretion (40). To determine which regions of PR play a critical role in the suppression of LHβ gene expression, transient transfection assays were performed using the PRA isoform and a PRB C557A DBD mutant receptor unable to bind to DNA. As observed previously, PRB suppressed basal LHβ promoter activity by 46% and the GnRH induction by 56%. The PRB-DBD mutant resulted in a complete disruption of R5020 repression in the absence or presence of GnRH induction (Fig. 2B). Similarly, when an expression vector for the mouse PRA isoform was transfected into L βT2 cells, there was no significant suppression of basal gene expression by progesterone (Fig. 2C). In the presence of PRα, suppression of GnRH induction was reduced by 19%, but the trend did not approach statistical significance. These data suggest that the PR DBD plays a critical role and that the unique amino-terminal region of PRB is involved in suppression of basal transcription and GnRH induction of the LHβ subunit.

**Mutation of Sp1, SF-1, or Egr-1 elements in the distal or proximal LHβ promoter does not alleviate progesterone suppression of LHβ transcription**

One potential mechanism for progesterone suppression of LHβ transcription is through an interaction between ligand-bound PR and the transcription factor Sp1 similar to the interaction reported between androgen receptor (AR) and Sp1 on the rat LHβ promoter (24). Alternatively, ligand-bound PR may interact with proximal promoter binding transcription factors, like the AR/SF-1 interaction observed on the bovine LHβ promoter (25).

To investigate the hypothesis that Sp1 binding sites are critical for PR suppression of LHβ transcription, specific mutations in Sp1 elements were created within the −1800-bp rat LHβ promoter. Figure 3A illustrates the location of the transcription factor binding sites on the LHβ promoter. The 5′ Sp1/CArG double mutation and the 3′ Sp1 mutation increased basal transcription compared with wild type (1.5- and 1.8-fold, respectively), whereas the triple mutation decreased basal activity by 93% (data not shown). In contrast, GnRH induction was not significantly altered by these mutations (Fig. 3B). There was no discernible difference in basal gene expression or GnRH induction of LHβ among the wild-type LHβ or these mutants after R5020 treatment. These data suggest that the Sp1-binding elements do not play a critical role in suppression of GnRH induction by progesterone.

Site-specific mutations were also generated in the 5′ SF-1, 3′ SF-1, 5′ Egr-1, and 3′ Egr-1 binding elements within the −1800-bp rat LHβ promoter. Each of these four mutations resulted in a reduction of basal promoter activity by approximately 40% (data not shown). GnRH treatment produced a 3.7-fold induction of the wild-type promoter (−1800 LHβ, Fig. 3C). Mutation of the 5′ SF-1 site led to a greater stimulation by GnRH, whereas mutation of the other proximal promoter binding sites resulted in a diminished response to GnRH: 3-fold with the 3′ SF-1 mutant, 1.6-fold with the 5′ Egr-1 mutant, and 1.5-fold with the 3′ Egr-1 mutant, similar to previous reports (8, 9, 43).
Regardless of the mutation, R5020 still repressed basal gene expression and suppressed the remaining GnRH induction, indicating that these proximal promoter elements are not essential for the suppression of LHβ gene expression by progestins.

**Progestosterone suppression of LHβ transcription maps to a region located −300/−150 upstream of the transcription start site of the LHβ promoter**

Because the suppression of LHβ gene expression by progesterone did not appear to require the known transcription factor binding elements, we used truncation analysis to identify the necessary regions. Truncation to −500, −150, or −87 resulted in significant reductions in basal activity in the absence of any hormones compared with the −1800-bp LHβ-luc reporter, whereas the −300 truncation increased basal gene expression (Fig. 4A). These results suggest that transcription factors important for basal gene expression also bind the −300/−150 region, in addition to the more distal and proximal regions of the LHβ promoter.

GnRH induction of the −500 LHβ truncation was comparable with that seen with the −1800-bp LHβ-luc reporter (Fig. 4B). However, the −500 LHβ truncation exhibited only a partial reduction by progesterone of basal promoter activity and GnRH induction compared with the −1800-bp LHβ-luc reporter, suggesting the presence of an upstream region involved in the suppression of GnRH induction of LHβ by progesterone. The −300 LHβ-luc reporter showed a significant repression of basal activity and GnRH induction after progesterone treatment (Fig. 4B). In contrast, the −150 LHβ truncation lost the suppression of basal gene expression and GnRH induction by progesterone, suggesting that the −300/−150 region of the LHβ promoter is
critical for the progesterone suppression. Interestingly, when the −87 LHB promoter was tested, the effects were reversed. RS020 treatment resulted in a significant increase in −87 LHB transcription (23.5%), and cells treated with both GnRH and RS020 showed a synergistic induction compared with the individual hormones.

The repressive element at −300/−150 of the LHB promoter is necessary and sufficient for progesterone suppression

Because the suppression mapped to the −300/−150 region using 5′ truncations of the LHB promoter, we tested whether this region was necessary for progesterone repression of both basal LHB gene expression and GnRH induction. Deletion of this region within the context of the −1800-bp LHB promoter still resulted in progesterone suppression (Fig. 4C). Given the results from the truncation analysis, we hypothesized that an upstream region may compensate for the repressive element at −300/−150, so we created a 150-bp deletion within the −500 LHB promoter. Basal gene expression was reduced by 40% (data not shown), indicating that this region has a role in LHB transcriptional regulation. Interestingly, there was no suppression of basal LHB gene expression or the GnRH induction by RS020 with the repressive element deleted in the context of the −500 LHB promoter (Fig. 4C), indicating that this 150-bp region (−300/−150) is necessary for the progesterone suppression in the context of the proximal promoter.

To assess whether the −300/−150 repressive element in the LHB promoter is sufficient for suppression by progesterone, we inserted the 150-bp element upstream of a heterologous TK-luc reporter in the forward or reverse orientation. Basal TK promoter activity was suppressed more than 50% by the addition of this element (data not shown). Progesterone further suppressed promoter activity by 30% when the −300/−150 repressive element was in the forward or reverse orientation (Fig. 4D), indicating that this 150-bp element from the proximal LHB promoter is sufficient to elicit the suppression by progesterone.

PR binding to the repressive element at −300/−150 is not critical for LHB suppression

Given that the −300/−150 region of the LHB promoter was essential for progesterone suppression, EMSA was performed to assess whether PR can bind to the proximal rat LHB promoter in vitro. Six 35-mer oligonucleotide probes were designed to span the −300/−150 region. Purified PR bound the −225/−191 oligonucleotide probe (Fig. 5A, lane 1), whereas no binding to the other probes in this region was observed (data not shown). To further demonstrate that PR bound specifically to the −225/−191 probe, the resulting complex was supershifted by a PR-specific antibody (lane 3) but not by IgG (lane 4). This complex also showed evidence of competition with a consensus PRE (data not shown). To further define where PR binds to the proximal LHB promoter, EMSA was performed with PR whole-cell extract using either wild-type (WT) −225/−191-labeled probe or oligonucleotides containing 3-bp scanning mutations as indicated, each with three adjacent A substitutions in the 3-bp sequence shown above each of the lanes. C, LβT2 cells were transiently transfected with either the −500 LHB-luc reporter plasmid or a reporter containing a PRE cis mutation in which the G and C residues important for high-affinity DNA binding were mutated (supplemental Table 1). Luciferase activity and set relative to the empty reporter vector. Results represent the mean ± SEM of at least three independent experiments performed in triplicate and are presented as fold induction of hormone treatment relative to the vehicle control. *, Significant differences from the vehicle-treated control; †, interaction as defined by a two-way ANOVA (P < 0.05).

FIG. 5. PR binds to a putative PRE at −221/−207, but mutation of the PRE does not alter progesterone suppression. A, Purified PRB was incubated with a wild-type (lane 1) or PRE mutant (lane 2) −225/−191 probe (supplemental Table 1) and tested for complex formation in EMSA. The addition of a PR antibody (PR Ab, lane 3) or nonspecific IgG control antibody (IgG, lane 4) to the binding reaction, are indicated. PR binding and the antibody supershift are also indicated. B, EMSA was performed with PR whole-cell extract using either wild-type (WT) −225/−191-labeled probe or oligonucleotides containing 3-bp scanning mutations as indicated, each with three adjacent A substitutions in the 3-bp sequence shown above each of the lanes. C, LβT2 cells were transiently transfected with either the −500 LHB-luc reporter plasmid or a reporter containing a PRE cis mutation in which the G and C residues important for high-affinity DNA binding were mutated (supplemental Table 1). Luciferase (Luc) activity was normalized to β-galactosidase activity and set relative to the empty reporter vector. Results represent the mean ± SEM of at least three independent experiments performed in triplicate and are presented as fold induction of hormone treatment relative to the vehicle control. *, Significant differences from the vehicle-treated control; †, interaction as defined by a two-way ANOVA (P < 0.05).
Two 50-bp regions of the repressive element are critical for LHβ suppression

Because direct binding of PR to the repressive region did not appear to be required for progesterone suppression, we created three sequential 50-bp deletions within the −500 LHβ promoter. Each of these three deletions resulted in greater than 50% reduction in basal gene expression (Fig. 6A), indicating that these regions must contribute to basal gene expression, most likely by binding specific transcriptional activators. The promoter containing the −250/−200 deletion was suppressed by progesterone for both basal LHβ expression and GnRH induction (Fig. 6B). In contrast, there was no significant progesterone suppression after either the −300/−250 or −200/−150 regions were deleted, indicating that more than one region of the 300-bp element is necessary for suppression by progesterone. In silico analysis of these 50-bp regions revealed a number of putative transcription factor binding elements. However, 10-bp scanning deletions through the two important regions did not prevent progesterone suppression of GnRH induction of the LHβ promoter (data not shown), suggesting that multiple elements within each region may be important.

Discussion

Our study demonstrates that progestins can suppress both basal transcription and GnRH induction of the LHβ gene in gonadotrope cells. Similar to androgens, progesterone suppression occurs in a hormone- and receptor-dependent manner, indicating that the actions of progesterone are through the classical PR. Moreover, the fact that PR is recruited to the endogenous LHβ promoter after hormone treatment and that the suppression occurred in as little as 6 h, indicate that progesterone mediates suppression of LHβ gene expression through a mechanism directly involving the LHβ promoter.

Maximal suppression of LHβ transcription by progesterone requires the hormone-bound PRB complex containing an intact DBD. Disruption of the PR DBD abrogated the repression of basal transcription as well as the suppression of GnRH induction by progesterone. One possible role for the PR DBD is for direct binding of PR to the LHβ promoter. To investigate this possibility, we searched for PREs within the −300/−150 repressive element necessary for progesterone suppression. We identified a putative PRE at −225/−191 that bound PR in EMSA. However, mutation of this site or deletion of a 50-bp region from −250/−200 that encompassed this element did not relieve progesterone suppression of LHβ transcription, suggesting that direct DNA binding by PR is not necessary for this effect, in contrast to progesterone induction of the FSHβ gene that requires direct binding of PR to the proximal promoter. Another possibility for the role of the PR DBD in the suppression of LHβ transcription by progesterone is through a protein-protein interaction with another transcription factor involved in LHβ expression, rather than direct DNA binding. For instance, Curtin et al. (24) demonstrated that the AR DBD interacts directly with Sp1 and as a result reduces Sp1 binding to the rat LHβ promoter, thereby decreasing the GnRH response. However, it is unlikely that Sp1 is involved in progesterone suppression because mutation of both of the Sp1 elements in the distal LHβ promoter had no effect and overexpression of Sp1 did not relieve repression (data not shown).

In addition to the PR DBD, we also demonstrated that the unique amino-terminal region of the PRB isoform is necessary for the full suppression of LHβ by progesterone. This region has been shown to contain a transactivation function that is thought to be the reason that PRB is generally a stronger activator than PRA. As a result of this additional region, PRB likely forms a more stable secondary and/or tertiary structure than PRA [recently reviewed by Bain et al. (44)] that favors the interaction of factor(s) necessary for progesterone suppression of LHβ gene expression.

Initially, we had hypothesized that the mechanism of action for progesterone suppression would be similar to the reported androgen suppression on either the rat (24) or bovine (25) promoter. Similarities between the two mechanisms include the necessity for liganded receptor and an intact DBD. The most significant difference is that the site of action for PR on the LHβ promoter is distinct from that of AR. Mutations in the distal Sp1 sites failed to relieve progesterone suppression of both basal and GnRH-induced LHβ gene expression, indicating that these ele-
ments are not critical for the suppression by progesterone. Mutation of the SF-1 or Egr-1 binding sites in the proximal promoter also did not relieve suppression by progesterone, suggesting that these elements do not play a critical role in progesterone suppression of LHβ mRNA levels. Our experiments with the 5′ truncations of the LHβ promoter provided supporting evidence for these conclusions. Specifically, a −300 LHβ-luc reporter lacking the distal Sp1 elements was sufficient for suppression by progesterone, and conversely, a −150 LHβ truncation (containing both Egr-1 and SF-1 binding sites) was not suppressed by progesterone.

In addition to highlighting the different mechanisms of repression by progestins vs. androgens, our analysis also revealed that several regions in the LHβ promoter contribute to the suppression by progesterone. Truncation analysis identified a repressive element between −300/−150 that was both necessary and sufficient to elicit progesterone suppression. Because this region is less conserved among mammalian species than the proximal LHβ promoter, it remains to be determined whether progesterone suppression of LHβ transcription occurs in a species-specific manner. The fact that deletion of the repressive element resulted in loss of suppression in the context of the −500 LHβ promoter but not the −1800-bp promoter suggests that a region upstream of −500 also contributes to progesterone suppression of both basal and GnRH-induced LHβ gene expression by progesterone.

To further characterize the critical region necessary for suppression of LHβ transcription in the proximal promoter, we created and tested three 50-bp deletions in the −300/−150 region. All three deletions reduced basal activity of the LHβ promoter by about 50%, indicating that they likely bind factors important for LHβ gene expression. Deletion of either the region from −300/−250 or −200/−150 resulted in a lack of suppression by progesterone, indicating that multiple elements may be required but that spacing is less critical because the middle 50 bp can be deleted without effects on progesterone regulation. Also supporting the idea that multiple elements in the proximal promoter may be responsible for the progesterone suppression is the fact that 10-bp scanning deletions through these two regions did not prevent progesterone suppression of GnRH induction. This situation is reminiscent of the mechanism of androgen suppression on the bovine LHβ promoter, in which cis mutations or block replacements affecting Egr-1, SF-1, or pituitary homeobox 1 binding elements had no effect, but the proximal promoter clearly mediated the androgen suppression (25).

Furthermore, the data thus far support the conclusion that the repression of basal activity and GnRH induction of the LHβ gene by PR occur through modulation of factor(s) in common between the two processes.

In summary, our results demonstrate that progesterone can suppress both basal transcription and GnRH induction of LHβ gene expression in a hormone- and receptor-dependent manner in gonadotrope cells. We determined that the full suppressive effect of progesterone on LHβ gene expression requires the unique amino-terminal region of the PRB isoform and an intact DBD. However, we did not find any evidence that the progesterone suppression involves direct binding of PR to the LHβ promoter, although it is recruited to the endogenous promoter in live cells. Rather, our data suggest that these domains are necessary for tethering or binding to other transcription factor(s).

Furthermore, we identified a repressive element in the proximal LHβ promoter that is both necessary and sufficient to elicit suppression by progesterone. Multiple regions at −300/−200 and −200/−150 appear to be involved in both basal transcription of the LHβ gene and suppression by progesterone, further supporting the concept that PR may act through other transcription factors bound to these regions. We also demonstrated that there is a region upstream of −500 in the rat LHβ promoter that may also be involved in the suppression of LHβ transcription by progesterone. Additional experiments will be necessary to define the cis-regulatory elements and transcription factors that play a role in the regulation of LHβ gene expression by ligand-bound PR. Altogether this work has revealed new insights into the pituitary action of progesterone. In particular, it has highlighted the role that progesterone may play in limiting preovulatory GnRH-induced LH secretion via progesterone suppression of GnRH-induced LHβ transcription.

Acknowledgments

The authors thank Djurdjica Coss and other members of the Mellon lab for helpful discussions and comments. We also thank Scott Kelley for his suggestions and critical reading of the manuscript. We are grateful to Mark Lawson, Benita Katzenellenbogen, and Dean Edwards for providing reagents. The −1800-bp rat LHβ luciferase-reporter plasmid (in pGL3) was donated by Mark Lawson. Rat PRB (in pCMV5) was provided by Benita Katzenellenbogen; the 1294 PR antibody and mouse PR and PRA plasmids (in pcDNA-I) were donated by Dean Edwards. We also acknowledge the University of California, San Diego, Cancer Center DNA Sequencing Shared Resource for sequencing and the University of Colorado Cancer Center Tissue Culture Core Facility for baculovirus production.

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This work was supported by National Institute of Child Health and Human Development/National Institutes of Health (NIH) through a cooperative agreement (U54 HD12303) as part of the Specialized Cooperative Centers Program in Reproduction and Infertility Research (to P.L.M.). This work was also supported by NIH Grant R01 HD20377 (to P.L.M.). V.G.T. was supported by NIH Grants F32 DK065437 and T32 HD07203.

Disclosure Summary: The authors have nothing to disclose.

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